

## GENETICS OF RETINITIS PIGMENTOSA – HISTORY REVISITED

Kalaivani. K <sup>1</sup>, Suganya Kannan <sup>2</sup>,  
Jeyakumar Balakrishnan <sup>3</sup> and Krishna Prasanth Baalann <sup>4\*</sup>

<sup>1,2</sup> Department of Ophthalmology, Vinayaka Mission's Medical College and Hospital, Vinayaka Mission's Research Foundation (Deemed to be University), Karaikal, Puducherry, India.

<sup>3</sup> Department of Central Research Laboratory for Biomedical Research, Vinayaka Mission's Medical College and Hospital, Vinayaka Mission's Research Foundation (Deemed to be University), Karaikal, Puducherry, India.

<sup>4</sup> Department of Community Medicine, Sree Balaji Medical College & Hospital, Bharath Institute of Higher Education & Research, Chennai, Tamilnadu, India.

\*Corresponding Author Email: mail2kristain@gmail.com

DOI: [10.5281/zenodo.12703159](https://doi.org/10.5281/zenodo.12703159)

### Abstract

Retinitis pigmentosa (RP) represents a heterogeneous group of inherited retinal disorders characterized by progressive degeneration of photoreceptor cells, leading to gradual loss of vision. Over the years, extensive research efforts have been directed towards unraveling the complex genetic landscape underlying RP. This review revisits the historical milestones in the understanding of RP genetics, tracing the evolution of knowledge from the initial identification of genetic loci to the recent advancements in next-generation sequencing technologies. The role of various genes implicated in RP pathogenesis, including those encoding proteins involved in phototransduction, structural integrity of photoreceptors, and cellular metabolism, will be discussed. Furthermore, insights into the genotype-phenotype correlations, inheritance patterns, and genetic modifiers influencing disease progression will be explored. The integration of molecular genetics with clinical phenotyping has facilitated the development of targeted therapies and personalized approaches for managing RP. Finally, future directions and challenges in deciphering the intricate genetic architecture of RP, as well as translating these findings into effective therapeutic interventions, will be highlighted. This review aims to provide a comprehensive overview of the genetic basis of RP, shedding light on its historical context and paving the way for further advancements in research and clinical practice.

**Keywords:** Retinitis Pigmentosa; Night Blindness; Gene Therapy; Mutations; Rhodopsin.

### INTRODUCTION AND GENETIC BASIS

The most prevalent inherited retinal degeneration is Retinitis Pigmentosa (RP). Night blindness, visual field constriction, and retinal pigmentation are the three main primary symptoms of RP. At the time, vitamin A insufficiency was more likely to be the cause of the night blindness described than RP (1). Before the ophthalmoscope was created, cadaver anatomic dissection was where retinitis pigmentosa was first identified. However, neither RP or night blindness were connected to it. Only with the invention of the modern ophthalmoscope can optometrist truly peep inside the lens of the eye and connect signs and symptoms such as blindness at night and limited field of vision to retina pigment (2).

Retinitis pigmentosa (RP), a degenerative illness that results in blindness, is brought on by inherited abnormalities in the rod visual pigment rhodopsin. Rhodopsin mutations account for more than 150 distinct mutations, and taken as a whole, this is a very prevalent root and causes a adRP (autosomal dominant RP) (1). Rhodopsin mutations are also linked to recessive RP (arRP) and dominant congenital stationary night blindness (adCSNB). Rhodopsin function loss typically characterizes recessive in RP, whereas rhodopsin function gain and/or dominant negative activity characterize dominant conditions. Here we classify rhodopsin mutations into seven types, with

abnormalities ranging from inappropriate localization, misfolding, and disruption of proteostasis to instability and altered function. A novel model for understanding how changes in photoreceptor homeostasis can result in neuronal cell death is provided from rhodopsin adRP (3).

### **Genetic Architecture and Inheritance Pattern of Retinitis Pigmentosa**

Rhodopsin (RHO) a gene mutation, will generate a rod photoreceptor retinal pigmentation, is the single most immediate cause of autosomal dominance (AD) RP. These mutations were the first genetic disorders discovered in retinitis pigmentosa (RP). G-protein-coupled receptor (GPCR) rhodopsin has been the subject of strong research in science. The most understanding function and degeneration of the retina caused by mutant rhodopsin. The functional phenotypic is not just one form of a disease. It has two primary phenotypic groups of RHO-ADRP, building on subclassification methods of ungenotyped ADRP (4).

From an early age, Class A mutants cause significantly strange in rod function and distributed throughout the retina, and cone density is connected with the residual cone function's topography. Even in adulthood, certain retinal areas or the entire retina of class B mutants can have practically normal rods, and there is a gradual characteristic illness process that involves an intraretinal gradient of disease susceptibility (5).

The cause and treatment of human RHO-ADRP remains unknown since postmortem donor retinas are rare and suffer from advanced disease stages (3). Without a doubt, the final shared mechanism of photoreceptor degradation in RHO-ADRP has been identified. Morphological associations were found between the T17M and P23H mutations and the intraretinal disease gradient in class B (6).

A sizeable majority of monogenic retinal diseases are X-linked retinopathies. They comprise both stable and progressive diseases, both with and without syndromic characteristics (2). Although many are X-linked recessive, a few display a characteristic in female carriers, which may help in diagnosis and provide details about disease causes. Carriers for an illness may falsely imply autosomal dominant inheritance. Some diseases, like RPGR-associated retinopathy, point out the limitations of existing genetic sequencing techniques by exhibiting various symptoms from variations in the same gene. Loss of function is a common cause of X-linked illness, suggesting that gene replacement procedures may be advantageous (7).

### **Genetic Variants and their Effects**

- Given the latest developments in next-generation sequencing, the molecular diagnostic is unclear affects between 30% and 40% of people with hereditary retinal disorders (IRDs)
- Variations in regulatory areas, intronic variants, and structural variants (SVs) (4).
- Depending on the population investigated, AdRP, which is responsible for 25%-40% of cases, was recently linked to 30 different gene alterations, most notably CA4 on Chr17q23.1.
- Whole-genome sequencing (WGS) was used to identify and characterize the complex SVs on Chr17q22 identified in the RP17 sense in several families as the genetic source of adRP
- The impact of RP17 SVs on the chromatin organization, regional epigenome, and

topologically associating domains (TADs) that compartmentalize the genome in three dimensions. Enhancer promoter interactions in the nuclear 3D space are made possible by TADs, which are chromatin domains inside the genome. When TAD structures are disrupted, regulatory areas and their target genes may lose chromosomal contact, or unique active sites may develop and highlighting ectopic interactions between legal boundaries, a modification in genetic expression that are harmful, as well as a new target gene. We show that ectopic enhancer-gene interactions at the RP17 locus are caused by changed TAD structure, which is compatible with a significant increase in activity (8).

- All genotypes show a significant phenotypic connection with relatively moderate illness, lowered vision sensitivity, nyctalopia, constricted field of vision with gradual development in line using adRP. A lot of people still hold good center sight in their sixth or seventh decade. People with UK-SV2 frequently have cystoid macular edema and foveal sparing. Based on a small sample size of patients, UK-SV6 (with a triplicated SV) could be linked to a more serious phenotype and earlier age of onset (9).

### Genomic Studies and New Genetic Variants Identification

Retinitis pigmentosa (RP) is a term used to describe a group of related inherited retinopathies with widely varying clinical outcomes, numerous known mutations, and multiple disease-causing genes. To find the genes and mutations causing these ailments, afflicted people and their families must undergo both gene discovery and mutation screening. This is essential to the development of medicines (10).

- NGS, which is often known as high levels of throughput or "deep sequencing," is a kind of next-generation DNA sequencing, is one of the most effective methods for genetic testing. Several novel RP genes have been discovered by NGS, although a sizable portion of cases that were previously and there are variations in genes known can cause retinal diseases, though they are not usually RP (11).
- The genetics of RD are extremely complicated, as shown by the huge quantity of genome information that have been accessible because of the discovery of the one associated with the initial mutant linked to RP on peoples (Histidine to a proline alteration in the Rhodopsin's amino acid position 23, described authored by Dryja et and co.). With over 200 genetic markers discovered, study of the genetic factors of RD nearly fifty percent of those diagnosed had the illness gene identified (12).
- In addition to RP, these genes also play a role in a wide range of other clinical conditions, including LCA, macular degeneration, and CORD. Both the genes located in an X chromosome (xIRP) and 20 genes for an arRP and adRP have each been identified as of this writing (9). Several of these genes are subject to mutations that might result in autosomal-dominant or autosomal-recessive forms, depending on the mutation (13).
- Mutations in multiple genes cause chronic kinds of USH or separated non-syndromic being deaf (PCDH15, MYO7A, CH23, USH1G and USH1C), RP (USH2A gene), as well as improvements with regard to the exact same gene causing various clinical entities, as has been noticed for ABCA4, which is involved in arRP, arMD, and arCORD. Furthermore, the majority of RP-causing mutations are unique to one or a few people or families (14).

- There are numerous proteins and genes that are linked to RD. The breakdown of protein in the pigment epithelium of the retina, ionic exchange, or the movement of compounds in the ribbon synapses of photoreceptors are just a few of the additional roles that these proteins can play in addition to their contribution to retinal actions (11).
- The proteins encoded by the known RP-associated genes perform a wide range of tasks, including pre-mRNA transcription (PRPF31; OMIM\_606419), transcriptional control of photoreceptor differentiating (NRL; OMIM\_162080), specific to rod-like structures phototransduction (RHO; OMIM\_180380), and the function of the cilia (C2orf71; OMIM\_613428). The most often occurring mutations among Asian (particularly Japanese) patients are EYS (OMIM\_612424) [12], RP1L1 (OMIM\_608581) [13], and CYP4V2 (OMIM\_608614), whereas the most frequently occurring genomes found in Caucasians RP examination are RHO] and USH2A (15).
- Although over 260 genes have so far been linked to IRDs, only roughly 80% of hereditary illnesses are thought to be caused by these genes (13). The development of next-generation sequencing, which combines molecular diagnosis, identification of genes for retinal disorders, and in silico functional evaluation, is essential for finding the remaining 20–30% of uncommon variants in the IRDs (16).
- The GPCR and olfactory receptor signaling pathway is activated by the protein-coding gene OR56A5 (Olfactory Receptor Family 56 Subfamily A Member 5). The conditions Nager Type, Acrofacial Dysostosis 1, and Acrofacial Dysostosis are linked to OR56A5. The understanding of smell signals and G-protein-mediated transduction are carried out by the body's olfactory receptors, which have a seven-transmembrane domain structure and a variety of neurotransmitter and hormone receptors. The largest gene family in the genome is the olfactory receptor gene family in particular. The second gene we identified is OR52L1 (Olfactory Receptor Family 52 Subfamily L Member 1), and it is a member of the same genetic family as pseudobulbar palsy. The first neurological reaction that results in the perception of smell is triggered by odorant molecules and olfactory receptors in the nose. (17).

### **Pathway And Mechanism Implicated**

The implications of apoptotic as well as non-apoptotic death of cells procedures during RP, as well as the different biological processes that initiate those channels in degrading photoreceptors, will be addressed in the section that follows.

### **Apoptosis**

The activation of caspase proteins is essential for the initiation of apoptosis. Intrinsic apoptosis can be induced by the elimination of the cells injury or the loss of survival indications, resulting in a higher level of proteins that promote apoptosis such as Bax and the release of proteins from mitochondria such as cytochrome c. Apoptotic Protease, which is Activated Factor I (APAF1) and caspase-9 (18) is created by cytoplasmic cytochrome c (18).

Caspase-9, which activated this complex, subsequently degrades and releases caspases that are executioners (caspase-3, 6, and 7). This starts off a series of events that leads to DNA and protein breakage, cytoskeleton condensation and chromatin, and the creation that leads to apoptotic bodies. Extrinsic apoptosis can also be

triggered by interactions among ligands manufactured by immune system cells and death receptors that are located on the surface of injured cells (for example, Tumors Necrosis Factor (TNF) or Fas). Ligand-bound receptors attract and activated caspase-8, which results in the stimulation of execution the caspases and the development of bodies that are apoptotic (19).

Both of these processes result in the oligomerization of CytC, APAF1, and pro-caspase-9 resulting in the apoptosome once CytC is released from the mitochondria. Several regulated necrotic processes can lead to photoreceptor death. Necroptosis demands RIP1/RIP3 activation, which stimulates and polymerizes MLKL to create holes in the membrane of the cytoplasm (20).

High cGMP levels can activate PARP, which may cause AIF to translocate into the center of the cell and result in parthanatos. Although the mechanism is unknown, certain RP-causing mutations could process to high Fe<sup>2+</sup> or GPX4 control, which would accelerate lipid oxidation and cause death by iron metabolism (21).

Endoplasmic reticulum (ER) anxiety can start macro autophagy by activation of ATG12, immediate stimulation of ATG18/WIP12, and action of beclin1. Lysosomal enzymes proceed to break down the interior of autophagosomes, which are formed when phagophores connect with lysosomes to create (22).

These include micro autophagy by expansion of the lysosomal membrane and chaperone-mediated autophagy with hsc-70 assist are able to transport proteins into lysosomes. The letters CytC, Glutathione peroxidase 4, Apoptotic Protease-activating Factor 1 (APAF1), MLKL (mixed lineage kinase domain-like pseudokinase), PARP (poly-ADP ribose polymerase), RIP (receptor-interacting serine/threonine-protein kinase), AIF (apoptosis-inducing factor), and TG (autophagy-related protein) are the various symbols for cytochrome C.

### **Regular necrosis**

Even though apoptosis has been described in retinal degeneration models, there has been increasing proof that caspase-independent death of cells processes additionally play a role in many blinding illnesses, notably RP. In fact, current study indicates that these alternate methods may be the primary methods of dying cells in RP (23).

Lately several types of controlled necrosis have been mentioned, such as necroptosis, ferroptosis, pyroptosis, and parthanatos. Each of the above processes result in a breakdown in the cytoplasmic membrane, because associated tissue damage causes an important inflammatory response. Different molecular mechanisms, however, play a role in their origin (24).

The coordination of two receptor binding protein kinases (RIP1 and RIP3) controls necroptosis. This association causes phosphorylation and polymerization of the mixed lineage kinase domain-like protein (MLKL), leading in the formation of the membrane spaces and the loss of the cell's outer stability. This process can be started by activation the death of cells harm to DNA sensors, toll-like receptors, interferons, and receptors (25).

The buildup of reactive oxygen species (ROS) and enhanced lipid peroxidation during ferroptosis, which is defined by an overproduction of intracellular iron (Fe<sup>2+</sup>) and dysregulation of glutamate metabolism, improve photoreceptor duration. This indicates in some types of RP; iron overload may also lead to cell death.



Pyroptosis is another kind of apoptotic which leads to cell membrane lysis and a response of inflammation, despite being dependent on caspase activation. When anti-inflammatory caspase-1 breaks down gasdermin D or gasdermin E, pyroptosis occurs. These proteins' N-termini enter into the cell membrane, producing permeabilization and rupture. Thioredoxin reduction led to the identification of pyroptosis in injured retinal pigment epithelium (RPE) cells, as there was no indication of photoreceptor pyroptosis in RP (26).

### **Autophagy**

Autophagy is a process in which components within cells are broken down and repurposed by lysosomes. The three main types of autophagy are chaperone-mediated autophagy (CMA), micro autophagy, and macro autophagy. When macro autophagy occurs, autophagosomes take up cytoplasmic material and merge with lysosomes to facilitate destruction. (27).

Consensus KFERQ proteins are directed toward lysosomes through the CMA-associated membrane glycoprotein 2A (LAMP-2A) receptors, where they are attached by the Hsc70 chaperone. During micro autophagy, the material that has to be destroyed enters lysosomes by invading the lysosomal or endosomal membrane.

Breakdown of injured cells and abnormally shaped or protein aggregates occurs by autophagy, which is essential. To maintain tissue and cell balance, the response of cells to oxidant and metabolic stresses may also induce autophagy. Autophagy, however, may end in the death of apoptotic cells once damage is severe (28).

### **Clinical And Genetic Heterogeneity**

As a large number of 93 RP families, clinical evidence of poor eyesight and blindness have been examined in connection to several mechanisms related to genetic transmission. Two clinical groups of autosomal-dominating RP might be recognized through the timing of macular involvement. While macular involvement took place after 20 years in the mild form, it was happened within 10 years in the severe form. Unexpectedly, a significant rise in mean paternal age (38.8 years vs. 29.1 years in controls in France, P less than 0.001) was detected in this variant of RP, indicating the presence of newly identified mutations. According to the age of initial and the sequence of development, four clinical subtypes of autosomal recessive RP might be differentiated (P less than 0.001), such as cone-rod dystrophy and early-onset serious cases (mean age of onset = 7.6 years), later-onset mild types, and aged forms (29). According to both the way and age of onset, two different clinical groups of X-linked RP have been recognized: whether myopic (mean age = 3.5 +/- 0.5 years) or blindness at night (mean age = 10.6 +/- 4.1 years). P lower than 0.001). The clinical development of the illness, however, which was extremely severe regardless of the clinical subgroup (blindness around the age of 25), showed little change. All these necessary carriers were researched and many studies shows pigmentation deposits in their peripheral retinas or severe eyesight. And many results show, 42% of the cases have series sporadic RP (30).

This group showed significant variation, and at minimum three diagnostic forms, namely cone-rod dystrophy, initial onset-severe forms, and advanced, mild types, could be detected. In the early stages of this illness, it was exceedingly challenging to determine whether the sporadic forms were inherited (particularly between 7 and 10

years), and the only thing that might have revealed the source of heredity was the clinical history. But strong similarity and a high gender ratio in the early stages and serious sporadic kinds (such as cone-rod dystrophy) indicated autosomal or X-linked recessive inheritance, but greater paternal age in advanced forms suggested dominant autosomal mutation (31).

### **Epigenetic Modifications and Their Role**

Several RPRPD-related genes control photoreceptor development and maturity in the human retina. Because the growth of the retina starts with a group of identical, expanding the retinal progenitor cells (RPCs) with particular "competence" for producing every kind of the retinal cells, which includes cone and rod photoreceptors, that we studied epigenetic changes in promoters of photoreceptor-related genes and gene sequences related to RPRPD at RPC differences towards cone and a rod photoreceptor (32).

Our findings indicate that, in comparison to other epigenetic markers or mechanisms, DNA methylation—a type of epigenetic sign—and DNA demethylation—a process—may be more important in the pathophysiology of these illnesses. Because genes with hypermethylated promoters in RPCs account for at least 40% of autosomal recessive RP cases and the majority of autosomal dominant RP cases, anomalies in the DNA demethylation mechanism during the RPC-to-photoreceptor transition may be important contributors to the cause and effect of retinitis pigmentosa (RP). According to the source, the extensive analysis pointed to an epigenetic model in which the failure to demethylate certain sequences (such as enhancers or promoters) of the genes required for photoreceptor growth, maturation, and function during the RPC-to-photoreceptor transition could reduce or eliminate their function, leading to RPRPD in the absence of any heritable genetic changes in these genes (33).

It's interesting to note that instances have been found in mouse and human RPCs, with noteworthy outcomes. It was discovered that 18 RP-related genes were hypermethylated. While some genetic promoters were hypomethylated in one species and hypermethylated in another, the majority of the hypermethylated genes were located in the RPCs of both species. The genes *CNGB1*, *IMPG1*, *IMPG2*, *NR2E3*, *PDE6A*, *PDE6G*, *PRPH2*, *RBP3*, *RHO*, *RP1*, *RPE65*, and *USH2A* are among those that have been linked. Notably, mutation changes in human *CNGB1*, *RHO*, *NR2E3*, *PDE6A*, *PRPH2*, *RP1*, *PDE6G*, *RPE65*, and human *EYS* (no homolog was found in mouse genomics) account for at least 40% of autosomal recessive RP cases (including *EYS*/10–20%, *USH2A*/17%) and autosomal dominant RP cases (30%, including *RHO*/25%). It is significant to note that several of these genes had "empty" states of chromatin in their hypermethylated promoters. It is to be expected that the majority of genes with hypermethylated promoters in mouse RPCs also have hypomethylated promoters in mouse rod and cone photoreceptors. Only two genes are now shown to be in a "bivalent" (temporarily repressed) chromatin state: *Tulp1* in murine RPCs and *LRAT* in human RPCs. These genes' promoters exhibited hypomethylation. 1% of human cases of autosomal recessive RP are linked to mutations in the *TULP1* and *LRAT* genes. In mature photoreceptors, the *Tulp1* promoter, which is "bivalent" in mouse RPCs, was in a receptive state of chromatin (34).

The study of CCRD-related genes showed that the promoters of six genes in mouse RPCs and 10 genes in human RPCs are hypermethylated. In both species, the

promoters of the following genes showed hypermethylation: AIPL1, KCNV2, PDE6C, PDE6H, and PRPH2. The only two genes shown to display "bivalent" chromatin states are CNGA3 in human RPCs and Guca1a in mouse RPCs. It's possible that a very small proportion of CCRD patients have mutations in these genes. The results of the previous investigation of the CSNB genes indicated that the promoters of seven genes in human RPCs and four genes in mouse RPCs were both hypermethylated. The promoters of the GNAT1, GRK1, and RHO genes were found to be hypermethylated in both species. In resistant chromatin states, no gene promoters were discovered. It is noteworthy that autosomal recessive CSNB is frequently caused by mutations in GNAT1 and RHO (32). Eight genes in human RPCs and six genes in mouse RPCs have hypermethylated promoters, according to a study on the genes associated with LCA that was published. There is evidence that human and mouse RPCs have hypermethylated promoters for AIPL1, DTHD1, RD3, RPE65, and PRPH2. Two genes are in a "bivalent" chromatin state, akin to RP, according to genetic research: Tulp1 in mouse RPCs and LRAT in human RPCs.

With the exception of RPE65, gene mutations might not be the cause of a sizable portion of LCA situations. A high incidence has been reported in the human (4 genes) and mouse (6 genes) RPCs had hypermethylated promoters for the genes connected to MD.

CFH, IMPG1, and PRPH2 are the genes whose promoters exhibited hypermethylation in both species. No repressive promoter-containing genes were found in mouse RPCs, in contrast to the single "bivalent" gene, PRDM13, which was found in human RPCs. It is important to remember that PRPH2 mutations account for 25% of Best vitelliform macular degeneration cases. Many promoters of genes related with CCRD, CSNB, LCA, and MD that were "bivalent" or hypermethylated in RPCs were cooperative and hypomethylated in developing cone or rod photoreceptors. We also discovered that many hypermethylated promoters had a "empty" chromatin state. (36).

### **Genotype -Phenotype Correlations**

As of right now, 284 PRPH2 variants—107 truncation variants, 149 missense variants, 21 in-frame variations, six gross elimination or insertion variants, and one 5' uncorrected variation have been reported in the literature. 32 nonsensical, 61 frameshift, 12 canonical splicing site alterations, 2 start loss variations, and 61 frameshifts are some of these variants. Based on published data, c.514C>T/p. R172G is the most often occurring variant of PRPH2, with a frequency of alleles of 11.6% (119/1028). This variant differs from the most common variant, p. G305Afs\*19, in our cohort. The most prevalent truncation mutation was c.828+3A>T, with an allele frequency of 6.6% (68/1028). While truncation variants were found across the coding frame, 119/149 (79.9%) of the missense alterations in PRPH2 were found in the peripherin protein's intradiscal D2 loop (ID2). (23). The 149 missense variants were examined utilizing in silico approaches, comparative validation with the gnomAD database, and final assessment in accordance with the ACMG/AMP criteria, as with the in-house discovered missense variants. All eighteen missense variants were thought to be benign or probably benign since they exhibited the following traits. (37).

Numerous alleles in the genome database; (2) considered benign by more than three prediction structures. (3) People with the same mutation and displayed unrelated symptoms (4) lack of co-segregation data in reported households.



The genotypic and phenotypic range of PRPH2 variations found in both sample and published research. (A) All discovered PRPH2 variations pathogenic or probably pathogenic PRPH2 variations in the cohort, the genome database, and known PRPH2 polymorphisms in the literature were dispersed and examined. Missense, truncation, and in-frame versions are shown by the blue, red, and gray lines, respectively. ID2 denotes the peripherin protein's intradiscal D2 loop. (B) In addition, the phenotypic range of the families with the detected PRPH2 variants is shown in the three pie charts. The cohort study has taken into consideration all non-Asian families as well as Asian families with PRPH2-associated retinopathy reported in the literature (38).

Although autosomal dominant inheritance was the most common kind, recessive autosomal dominant and digenic forms with a mutation of the homologous ROM1 gene were also found, but they were rather infrequent. This is according to reports on the different roles and mechanisms in vivo and in vitro examinations of rod and cone maintenance. A broad variety of rod- and cone-dominant instances of retinal aging are thought to be caused by pathogenic deletions of PRPH2, characterized by substantial inter- and intrafamilial particular heterogeneity. Based on studies using models with causative PRPH2 distinctions. Haploinsufficiency-related loss-of-function mutations, including p.C214S and p.P21 6L, contributed to the rod-dominant retinal disorder. Cone-dominant phenotypes were produced by the Gain-of-function alterations (p.C150S and p.R172W) (39).

However, PRPH2-associated retinopathy's genotype-phenotype relation remained unknown, yet in clinical settings, the identification, prognosis, and genetic counseling of PRPH2 variations remain challenging, especially for ophthalmologists who have little experience with this gene. More than 75% of patients with missense PRPH2 mutations, which are not present in the general population's database and may be hazardous by at least six of the seven computational approaches, suffered from RP (40).

### **Animal Models and their Contributions to Understanding Retinitis Pigmentosa Genetics**

To look into possible treatments that could extend photoreceptor lifetime and further investigate the mechanisms behind photoreceptor degeneration, multiple animal models were developed.

The outer retina's oxygen levels rise following severe rod mortality. Cone photoreceptors in Guinea experience constant oxidative stress due to rod breakdown, leading to a significant degree of cone degeneration, according to a role-playing model. Oxidative stress causes cell cone death because it maintains cone function and promotes cone survival. Retinal hyperoxia was assumed to be the first event that initiates the generation of reactive oxygen species, which in turn initiates the oxidative stress process in the RP retina. This, however, must be shown. (37).

Rod photoreceptors are the major focus of RP, a genetically heterogeneous disease that causes both initial rod death and secondary cone loss. After rod photoreceptor mortality, cone photoreceptors in RP undergo hyperoxia as a result of an excessive number of unnecessary oxygens. As a consequence, cones undergo oxidative stress, resulting leads to structural and functional damage and eventually leads to the cone's death. Approximately, research demonstrates that reducing oxidative stress on cones by exposing them to hypoxia recovers cone structure as well as function (41).

## Current and Future Prospects for Therapeutic Approaches Based on Genetic Insights

The dominant inheritance of the mutant gene makes genetic therapy for autosomal dominant retinitis pigmentosa (adRP) challenging. Treatment for adRP appears to require a combination of wild-type replacement of the key gene and mutant reduction. By administering a full-length human or mouse genomic rhodopsin gene using nanoparticles (NP), one of the dominant negative effects of the mutant rhodopsin might be offset in a medicinally meaningful way to P23H+/-knock-in heterozygous mice. The reported findings show mouse in the gRho and gRHO NP-treated groups also revealed important both structural and functional improvement of the rod photoreceptors, and this rehabilitation lasted for a period of time following injection, suggesting a prospective reduction in photoreceptor degradation. In order to find miRNAs that may be employed as rhodopsin gene enhancers or suppressor genes for sustained photoreceptor rescue, we did miRNA analysis of the transcriptome using next-generation genome sequencing and identified differently regulated miRNAs. It represents and shows the highlight of significance in the gene augmenting and utilizing a gDNA which contains regulating elements, showing that delivering a complete when contrasted with using cDNA for treating of this kind of adRP, utilising genetic region to generate transgenic had a higher probability of success.

Variations that affect the rhodopsin genes (RHO) generate around 40% of circumstances with autosomal-dominating RP (adRP). A reasonable hope is that in a novel cause of adRP, were characterized the clinical features in RHO's initial mono-allelic copy number variations (CNV).

- (1) It demonstrates a male patient in his 60's with extensive retinal degeneration and four transcriptionally active intact versions of rhodopsin
- (2) Retinal organoids are used to summarize the clinical phenotypes.
- (3) Analyze the clinical viability of using the small molecule Photoregulin3 (PR3) to concentrate on or limit progression of illness in RHO-CNV RP subjects.

Rhodopsin protein (RHO) expression was elevated and mis localized within the rod photoreceptor cell body by western blotting and immunohistochemistry in patient retinal organoids. These findings were confirmed by microscopy, increased RHO mRNA levels, and immunohistochemistry. Last but not least, the reported cases were associated with altered RHO expression by focusing on NR2E3, in the upstream regulator of RHO. Results mainly corrected RHO protein relocation within the midline to the inner/outer regions of rod photoreceptors also indicated the possibility of developed therapies. We truly need large databases that include precise genomic and clinical data in order to do the thorough genotype-phenotype comparisons. This may have predictive significance for the chance that the ailment will manifest and treatment alternatives, as well as critical information needed to identify RP in a patient or family for genetic counseling.

## CONCLUSION

In summary, the genetics of retinitis pigmentosa (RP) has evolved significantly, from initial genetic loci discovery to the current era of high-throughput sequencing. Decades of research have identified numerous RP-associated genes, providing crucial insights into the complex molecular mechanisms underlying the disorder. Understanding

genotype-phenotype correlations has facilitated genetic counseling and personalized management strategies. Integration of molecular genetics with clinical phenotyping has paved the way for targeted therapies and gene-based interventions. Challenges remain in translating genetic findings into effective treatments, highlighting the need for further exploration of genetic modifiers and environmental factors influencing RP pathogenesis. Collaboration across disciplines is essential for advancing our understanding of RP genetics and translating discoveries into clinical practice. By leveraging historical knowledge and embracing emerging technologies, personalized precision medicine approaches offer hope for improving outcomes for individuals affected by RP.

### Acknowledgement

The authors gratefully acknowledge VMRF-DU for financial assistance in the form of seed money. Aug 2021/VMRF/Seed Money/VMMC-Karaikal/3

### Conflict of Interest

There are no conflicts of interest among the authors.

### References

- 1) Wang HH, Chen N, Wang NK. Introduction and Discovery of Retinitis Pigmentosa. In Retinitis Pigmentosa 2022 Dec 9 (pp. 1-14). New York, NY: Springer US.
- 2) Marmor MF, Aguirre G, Arden G, Berson E, Birch DG, Boughman JA, Carr R, Chatrian GE, Del Monte M, Dowling J, Enoch J. Retinitis pigmentosa: a symposium on terminology and methods of examination. *Ophthalmology*. 1983 Feb 1;90(2):126-31.
- 3) Bell J. The Treasury of Human Inheritance: Nettleship Memorial Volume. Anomalies and Diseases of the Eye. Retinitis Pigmentosa and Allied Diseases. Cambridge University Press; 1922.
- 4) Athanasiou D, Aguila M, Bellingham J, Li W, McCulley C, Reeves PJ, Cheetham ME. The molecular and cellular basis of rhodopsin retinitis pigmentosa reveals potential strategies for therapy. *Progress in retinal and eye research*. 2018 Jan 1; 62:1-23.
- 5) Andrés A, Garriga P, Manyosa J. Altered functionality in rhodopsin point mutants associated with retinitis pigmentosa. *Biochemical and biophysical research communications*. 2003 Mar 28;303(1):294-301.
- 6) Kanan Y, Hackett SF, Hsueh HT, Khan M, Ensign LM, Campochiaro PA. Reduced inspired oxygen decreases retinal superoxide radicals and promotes cone function and survival in a model of retinitis pigmentosa. *Free Radical Biology and Medicine*. 2023 Mar 1; 198:118-22.
- 7) Zhang H, Wang A, Li G, Zhai Q, Huang Z, Wang X, Cao Z, Liu L, Liu G, Chen B, Zhu K. Osteoporotic bone loss from excess iron accumulation is driven by NOX4-triggered ferroptosis in osteoblasts. *Free Radical Biology and Medicine*. 2023 Mar 1; 198:123-36.
- 8) Hartong DT, Berson EL, Dryja TP. Retinitis pigmentosa. *The Lancet*. 2006 Nov 18;368(9549):1795-809.
- 9) Wang Y, Wang J, Jiang Y, Zhu D, Ouyang J, Yi Z, Li S, Jia X, Xiao X, Sun W, Wang P. New Insight into the Genotype-Phenotype Correlation of PRPH2-Related Diseases Based on a Large Chinese Cohort and Literature Review. *International Journal of Molecular Sciences*. 2023 Apr 4;24(7): 6728..
- 10) Arikawa K, Molday LL, Molday RS, Williams DS. Localization of peripherin/rds in the disk membranes of cone and rod photoreceptors: relationship to disk membrane morphogenesis and retinal degeneration. *The Journal of cell biology*. 1992 Feb 1;116(3):659-67.
- 11) Molday RS, Hicks D, Molday L. Peripherin. A rim-specific membrane protein of rod outer segment discs. *Investigative ophthalmology & visual science*. 1987 Jan 1;28(1):50-61.

- 12) Gamundi MJ, Hernan I, Muntanyola M, Trujillo MJ, García-Sandoval B, Ayuso C, Baiget M, Carballo M. High prevalence of mutations in peripherin/RDS in autosomal dominant macular dystrophies in a Spanish population. *Molecular Vision*. 2007; 13:1031.
- 13) Chen Y, Zhang Q, Shen T, Xiao X, Li S, Guan L, Zhang J, Zhu Z, Yin Y, Wang P, Guo X. Comprehensive mutation analysis by whole-exome sequencing in 41 Chinese families with Leber congenital amaurosis. *Investigative ophthalmology & visual science*. 2013 Jun 1;54(6):4351-7.
- 14) de Bruijn SE, Fiorentino A, Ottaviani D, Fanucchi S, Melo US, Corral-Serrano JC, Mulders T, Georgiou M, Rivolta C, Pontikos N, Arno G. Structural variants create new topological-associated domains and ectopic retinal enhancer-gene contact in dominant retinitis pigmentosa. *The American Journal of Human Genetics*. 2020 Nov 5;107(5):802-14.
- 15) Lidgerwood GE, Morris AJ, Conquest A, Daniszewski M, Rooney LA, Lim SY, Hernández D, Liang HH, Allen P, Connell PP, Guymer RH. Role of lysophosphatidic acid in the retinal pigment epithelium and photoreceptors. *Biochimica et Biophysica Acta (BBA)-Molecular and Cell Biology of Lipids*. 2018 Jul 1;1863(7):750-61.
- 16) Golovleva I, Köhn L, Burstedt M, Daiger S, Sandgren O. Mutation spectra in autosomal dominant and recessive retinitis pigmentosa in northern Sweden. *Retinal Degenerative Diseases: Laboratory and Therapeutic Investigations*. 2010:255-62.
- 17) Cherry TJ, Yang MG, Harmin DA, Tao P, Timms AE, Bauwens M, Allikmets R, Jones EM, Chen R, De Baere E, Greenberg ME. Mapping the cis-regulatory architecture of the human retina reveals noncoding genetic variation in disease. *Proceedings of the National Academy of Sciences*. 2020 Apr 21;117(16):9001-12.
- 18) Ibrahim DM, Mundlos S. Three-dimensional chromatin in disease: what holds us together and what drives us apart? *Current Opinion in Cell Biology*. 2020 Jun 1; 64:1-9.
- 19) Lin TY, Chang YC, Hsiao YJ, Chien Y, Jheng YC, Wu JR, Ching LJ, Hwang DK, Hsu CC, Lin TC, Chou YB. Identification of novel genomic-variant patterns of OR56A5, OR52L1, and CTSD in retinitis pigmentosa patients by whole-exome sequencing. *International Journal of Molecular Sciences*. 2021 May 25;22(11):5594.
- 20) Albarry MA, Hashmi JA, Alreheli AQ, Albalawi AM, Khan B, Ramzan K, Basit S. Novel homozygous loss-of-function mutations in RP1 and RP1L1 genes in retinitis pigmentosa patients. *Ophthalmic Genetics*. 2019 Nov 2;40(6):507-13.
- 21) Tsai FJ, Lee YC, Chang JS, Huang LM, Huang FY, Chiu NC, Chen MR, Chi H, Lee YJ, Chang LC, Liu YM. Identification of novel susceptibility Loci for kawasaki disease in a Han chinese population by a genome-wide association study. *PloS one*. 2011 Feb 4;6(2): e16853.
- 22) Krämer A, Green J, Pollard Jr J, Tugendreich S. Causal analysis approaches in ingenuity pathway analysis. *Bioinformatics*. 2014 Feb 15;30(4):523-30.
- 23) Hunter DJ, Altshuler D, Rader DJ. From Darwin's finches to canaries in the coal mine—mining the genome for new biology. *New England Journal of Medicine*. 2008 Jun 26;358(26):2760-3.
- 24) Plenge RM, Scolnick EM, Altshuler D. Validating therapeutic targets through human genetics. *Nature reviews Drug discovery*. 2013 Aug;12(8):581-94.
- 25) Gao FJ, Li JK, Chen H, Hu FY, Zhang SH, Qi YH, Xu P, Wang DD, Wang LS, Chang Q, Zhang YJ. Genetic and clinical findings in a large cohort of Chinese patients with suspected retinitis pigmentosa. *Ophthalmology*. 2019 Nov 1;126(11):1549-56.
- 26) Aleman TS, Cideciyan AV, Sumaroka A, Windsor EA, Herrera W, White DA, Kaushal S, Naidu A, Roman AJ, Schwartz SB, Stone EM. Retinal laminar architecture in human retinitis pigmentosa caused by Rhodopsin gene mutations. *Investigative ophthalmology & visual science*. 2008 Apr 1;49(4):1580-90.
- 27) Jacobson SG, Cideciyan AV, Sumaroka A, Aleman TS, Schwartz SB, Windsor EA, Roman AJ, Stone EM, MacDonald IM. Remodeling of the human retina in choroideremia: rab escort protein 1 (REP-1) mutations. *Investigative ophthalmology & visual science*. 2006 Sep 1;47(9):4113-20.

- 28) Banin E, Cideciyan AV, Alemán TS, Petters RM, Wong F, Milam AH, Jacobson SG. Retinal Rod Photoreceptor–Specific Gene Mutation Perturbs Cone Pathway Development. *Neuron*. 1999 Jul 1;23(3):549-57.
- 29) Cideciyan AV, Aleman TS, Jacobson SG, Khanna H, Sumaroka A, Aguirre GK, Schwartz SB, Windsor EA, He S, Chang B, Stone EM. Centrosomal-ciliary gene CEP290/NPHP6 mutations result in blindness with unexpected sparing of photoreceptors and visual brain: implications for therapy of Leber congenital amaurosis. *Human mutation*. 2007 Nov;28(11):1074-83.
- 30) Fishman GA, Stone EM, Gilbert LD, Kenna P, Sheffield VC. Ocular findings associated with a rhodopsin gene codon 58 transversion mutation in autosomal dominant retinitis pigmentosa. *Archives of Ophthalmology*. 1991 Oct 1;109(10):1387-93.
- 31) Boon CJ, van Schooneveld MJ, den Hollander AI, van Lith-Verhoeven JJ, Zonneveld-Vrieling MN, Theelen T, Cremers FP, Hoyng CB, Klevering BJ. Mutations in the peripherin/RDS gene are an important cause of multifocal pattern dystrophy simulating STGD1/fundus flavimaculatus. *British Journal of Ophthalmology*. 2007 Nov 1;91(11):1504-11.
- 32) Ngcungcu T, Oti M, Sitek JC, Haukanes BI, Linghu B, Bruccoleri R, Stokowy T, Oakeley EJ, Yang F, Zhu J, Sultan M. Duplicated enhancer region increases expression of CTSB and segregates with keratolytic winter erythema in South African and Norwegian families. *The American Journal of Human Genetics*. 2017 May 4;100(5):737-50.
- 33) Simell O, Takki K. Raised plasma-ornithine and gyrate atrophy of the choroid and retina. *The Lancet*. 1973 May 12;301(7811):1031-3.
- 34) Karpe G, Kornerup T, Wulff B. The clinical electroretinogram: VIII. The electroretinogram in diabetic retinopathy. *Acta Ophthalmologica*. 1958 Apr;36(2):281-91.
- 35) GOURAS P, CARR RE. Electrophysiological studies in early retinitis pigmentosa. *Archives of Ophthalmology*. 1964 Jul 1;72(1):104-10.
- 36) BIRD AC. Clinical investigation of retinitis pigmentosa. *Australian and New Zealand Journal of Ophthalmology*. 1988 Aug;16(3):189-98.
- 37) Zelante L, Gasparini P, Estivill X, Melchionda S, D'Agruma L, Govea N, Milá M, Della Monica M, Lutfi J, Shohat M, Mansfield E. Connexin26 mutations associated with the most common form of non-syndromic neurosensory autosomal recessive deafness (DFNB1) in Mediterraneans. *Human molecular genetics*. 1997 Sep 1;6(9):1605-9.
- 38) Farjo R, Naash MI. The role of Rds in outer segment morphogenesis and human retinal disease. *Ophthalmic genetics*. 2006 Jan 1;27(4):117-22.
- 39) Schwarz JM, Rödelberger C, Schuelke M, Seelow D. MutationTaster evaluates disease-causing potential of sequence alterations. *Nature methods*. 2010 Aug;7(8):575-6.
- 40) Eichler EE, Flint J, Gibson G, Kong A, Leal SM, Moore JH, Nadeau JH. Missing heritability and strategies for finding the underlying causes of complex disease. *Nature reviews genetics*. 2010 Jun;11(6):446-50.
- 41) Kobayashi K, Knowles MR, Boucher RC, O'Brien WE, Beaudet AL. Benign missense variations in the cystic fibrosis gene. *American journal of human genetics*. 1990 Oct;47(4):611.